PRADIMICINS A, B AND C: NEW ANTIFUNGAL ANTIBIOTICS II. *IN VITRO* AND *IN VIVO* BIOLOGICAL ACTIVITIES

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Pradimicins A, B and C specify novel antibiotics produced by Actinomadura hibisca No. P157-2 (ATCC 53557) possessing potent and broad antifungal activity in vivo. They showed moderate in vitro antifungal activity against a wide variety of fungi and yeasts including clinically important pathogens, and were highly effective in systemic infection with Candida albicans in mice after iv and im administrations. Pradimicin A showed in vivo therapeutic activity against C. albicans, Cryptococcus neoformans and Aspergillus fumigatus in both normal and immunocompromized mice. 5-Fluorocytosine- and azole-resistant C. albicans strains were susceptible to pradimicin A. This antibiotic also demonstrated therapeutic efficacy against lung candidiasis and aspergillosis, vaginal candidiasis and skin Trichophyton mentagrophytes infection in mice with iv or topical treatment. The LD₅₀ values after a single iv or im administration were 120 mg/kg and more than 400 mg/kg, respectively. Against various cultured mammalian cells, pradimicin A was noncytotoxic at 100 or 500 μ g/ml, and showed potent anti-influenza virus activity with an IC₅₀ value of 6.8 μ g/ml.

In the course of antifungal screening, pradimicins A, B and C were found in the culture of Actinomadura hibisca No. P157-2 (ATCC 53557)^{1~4}) and their chemical structures were determined to be a benzo[a]naphthacenequinone substituted by a D-alanine and sugars^{4~7}). This paper describes their *in vitro* and *in vivo* antifungal activities. Pradimicin A has been extensively evaluated for *in vivo* therapeutic activity in a variety of murine systemic and local fungal infections, for acute toxicity in mice and for other biological activities.

Materials and Methods

Antifungal Agents

Pradimicins A, B and C were prepared at Bristol-Myers Research Institute Co., Tokyo and used after dissolving in saline containing 10% DMSO and adjusting to pH 7.5 with 1.0 N NaOH. Amphotericin B (Fungizone, sodium deoxycholate complex) was purchased from Squibb, Japan and used by dissolving in saline for both *in vitro* and *in vivo* experiments. Ketoconazole was obtained from Janssen Pharmaceuticals and used as a suspension in saline containing 10% DMSO. 5-Fluorocytosine purchased from Sigma Chemical Company was dissolved in saline.

In Vitro Antifungal Activity

The MICs of the antifungal agents were determined by an agar dilution method on yeast morphology agar adjusted to pH 7.0 with 1/15 m phosphate buffer. A 5- μ l aliquot of fungal suspension containing 10^6 cells/ml, except for *Trichophyton mentagrophytes* No. 4329 which contained 10^7 cells/ml, was inoculated onto the surface of the antibiotic-containing agar plates with a multiinoculator. After incubation at 28°C for 40 hours, the lowest concentration of antibiotic causing virtually complete inhibition of fungal growth

[†] Deceased.

(MIC) was determined.

In Vivo Antifungal Activity

Systemic Fungal Infections in Normal Mice: The *in vivo* therapeutic efficacy of pradimicins A, B and C was examined comparatively with amphotericin B and ketoconazole against systemic infection with C. *albicans* A9540 (10⁶ cells/mouse), 5-fluorocytosine-resistant C. *albicans* YA22851 (5.5×10^5 cells/mouse), azole-resistant C. *albicans* YA26090 (8.8×10^5 cells/mouse), Cryptococcus neoformans IAM 4514 (10⁶ cells/mouse) or Aspergillus fumigatus IAM 2034 (10⁷ cells/mouse) in normal mice. C. *albicans* and C. *neoformans* were cultured at 28°C for 18 and 48 hours, respectively, in YGP medium (glucose 1.5%, peptone 1.0%, yeast extract 0.4%, K₂HPO₄ 0.05% and MgSO₄ · 7H₂O 0.05%, pH 7.0) and suspended in saline. A. *fumigatus* was cultured at 28°C for 7 days on YGP agar slant and the spores were suspended in saline. Male ICR mice weighing 20 to 24g were infected iv with approximately 10 times the median lethal doses of each fungus as shown above. Groups of 5 mice at each dose level were given the test compounds by iv or im dosing route. The initial treatment was given immediately after the fungal challenge. The 50% protection dose (PD₅₀) was calculated by the method of LITCHFIELD and WILCOXON⁸) from the survival rate recorded 20 days after the fungal challenge. Untreated animals died 7 to 15 days post infection.

Systemic Fungal Infections in Immunocompromized Mice: Groups of 5 male ICR mice weighing 20 to 24 g were treated ip with 200 mg/kg of cyclophosphamide 4 days before *C. albicans* A9540, *Candida tropicalis* CS-07, *C. neoformans* IAM 4514 or *A. fumigatus* IAM 2034 iv infection at 2.2×10^5 , 2.3×10^6 , 2.9×10^5 or 2.0×10^6 cells/mouse (10 times LD₅₀), respectively. Test compounds were administered iv or im to the mice. The initial treatment was done immediately after the fungal challenge.

Lung C. albicans and A. fumigatus Infections: Groups of 4 male ICR mice weighing 20 to 25 g were treated ip with 200 mg/kg of cyclophosphamide 4 days before the lung infection with C. albicans A9540 or A. fumigatus IAM 2034. The mice were anesthetized with ether and were infected with a 70- μ l of C. albicans or A. fumigatus cell suspension containing 1×10^6 or 1.1×10^8 cells, respectively, through an intranasal route on day 0. Test compounds were administered iv to the mice immediately after the inoculation followed by once a day administration on days 2 and 4. The PD₅₀ was calculated from the survival rate recorded 20 days after the fungal challenge.

Vaginal C. albicans Infection: Groups of 5 female ICR mice weighing 20 to 24 g were treated sc with 0.5 mg/kg of estradiol benzoate 3 days before and 2 days after vaginal infection with C. albicans A9540. A 10- μ l portion of a cell suspension of C. albicans containing 10⁶ cells was inoculated intravaginally on day 0. Test compounds were administered im to the mice immediately after the inoculation followed by once a day for 4 days. On day 7, vaginal exudate was sampled with a thin glass rod and spread on an YGP agar plate containing 100 μ g/ml of chloramphenicol. The plate was incubated at 28°C for 2 days. Viable cell count was made and graded by the scores indicated in Table 1.

Skin *T. mentagrophytes* Infection: Groups of 5 male ICR mice weighing 20 to 25 g were anesthetized ip with 35 mg/kg of pentobarbital. After removing the hair from the back (approx 2 cm^2), the skin was gently injured with sandpaper and vaccination needles. A 100-µl sample of cell suspension of *T. mentagrophytes* No. 4329 containing 5×10^5 cells was applied to the injured skin on day 0. Test compounds were mixed with Solbase (50% macrogol 400, Dainippon Pharmaceutical Co.) and topically spread around the infected area with a glass rod (once a day on days 0 to 4). On the day 7, lesions of the infected area were graded according to the severity of the infection by the scores indicated in Table 2.

Table 1.	Scores of vaginal	Candida	albicans	infection	ir
mice.					

Table 2. Lesion scores of skin *Trichophyton menta*grophytes infection in mice.

Number of cells/plate	Score	Severity of infection	Score
>459	4	Erythema and confluent scaling and	4
458~146	3	whitish patches	
145~47	2	Erythema and numerous whitish area	3
46~15	1	Erythema with whitish area	2
14~0	0	Erythema	1
		Normal	0

In Vitro Cytotoxicity

Madin Darby canine kidney (MDCK), green monkey kidney (Vero) and murine melanoma (B16-F10) cells were grown to the logarithmic phase in Eagle minimum essential medium (MEM) supplemented with fetal calf serum (FCS, 10% in MDCK and B16-F10 cells, and 5% in Vero cells) and antibiotic (50 μ l/ml of amikacin in MDCK and Vero cells, and 60 μ g/ml of kanamycin in B16-F10 cells). Human colon carcinoma (HCT-116) cells were grown in McCov's 5A medium supplemented with FCS (10%), benzylpenicillin (100 μ /ml) and streptomycin (100 μ g/ml) at 37°C in a humidified atmosphere of 5% CO₂ and 95% air. MDCK cells were obtained from the American Type Culture Collection (Rockville, MD) and the other cell lines were from Bristol-Myers Squibb Company (Wallingford, CT). MDCK, Vero, B16-F10 and HCT-116 cells were harvested and a 180- μ l aliquot of cell suspension was implanted into wells of a 96-well microplate at the inoculum sizes of 1×10^5 , 1×10^5 , 3×10^4 and 6×10^4 cells/ml, respectively, and incubated for 24 hours. After adding test compounds (20 μ l), they were further incubated for 72 hours. The cytotoxicity was colorimetrically determined at 540 nm after staining viable cells with 0.006% neutral red solution.

Acute Toxicity in Mice

Pradimicin A was dissolved in saline containing 10% DMSO. The test compound solution was adjusted to pH 7.5 with 1.0 N NaOH and injected iv or im once to groups of 5 male ICR mice at a dose volume of 0.2 ml per 10 g body weight. The animals were observed daily for 10 days for physical and behavioral signs of toxicity.

In Vitro Antiviral Activity Determined by the Plaque Reduction Assay

MDCK and Vero cells were grown in Eagle MEM supplemented with FCS (10% for MDCK cells, 5% for Vero cells) and 50 μ g/ml of amikacin. Influenza virus type A Victoria and herpes simplex virus type 1 (HSV-1) KOS strains were grown on MDCK and Vero cells, respectively. In the anti-influenza virus assay, confluent MDCK cell monolayers were grown in a 12-well microplate and the medium drained from the culture and washed with Eagle MEM. The culture was infected with $20 \sim 30$ pfu/well of the virus in Eagle MEM containing trypsin. After adsorption for 1 hour at 37°C, unadsorbed viruses were removed and 2 ml of an overlay of Eagle medium with 0.25% agarose containing 2-fold dilutions of test compound were added. The culture was incubated at 37°C in a humidified atmosphere of 5% CO₂ and 95% air for $48 \sim 72$ hours until the plaques were observed. After removing the overlay medium, cell monolayers were fixed with 10% formaldehyde solution and the plaques were counted. In the anti-HSV assay, Vero cell monolayers were grown in a 24-well microplate and the medium was drained from the culture, which was washed with Eagle MEM. The cultures were inoculated with approximately 50 pfu/well of HSV-1. After adsorption for 1 hour at 37°C, unadsorbed viruses were removed and the cultures were overlayed with 1 ml of Eagle MEM containing 0.3% immune serum globulin and 2-fold dilutions of the test compound. The cultures were incubated at 37° C for $48 \sim 72$ hours until the plaques were observed. After removing overlay medium, cell monolayers were stained with carbol-fuchsin and the plaques were counted.

Results

In Vitro Antifungal Activity

MICs for pradimicins A, B and C, amphotericin B and ketoconazole against 16 different fungi and yeasts are shown in Table 3. Pradimicin A showed weaker activity than amphotericin B but inhibited a wide range of fungi and yeasts tested with MIC values of $0.8 \sim 12.5 \,\mu$ g/ml except for *Mucor spinosus* which showed no susceptibility at 100 μ g/ml. Both pradimicins B and C exhibited similarly broad antifungal spectra while no or weak inhibition was seen against *Fusarium moniliforme*, *Petriellidium boydii* or *M. spinosus*. All pradimicins possessed significantly stronger anti-*Candida* activity than ketoconazole.

In Vivo Antifungal Activity against Systemic Infections in Mice

In vivo anti-Candida activity of pradimicins A, B and C was examined in the systemic C. albicans

			MIC (µg/m	1)	
Test organism	Pradimicins			A	¥
	Α	В	С	B B	Ketoconazole
Saccharomyces cerevisiae ATCC 9763	3.1	0.8	3.1	0.2	< 0.05
Candida albicans IAM 4888	6.3	3.1	3.1	0.4	50
C. albicans A9540	6.3	3.1	3.1	0.4	50
C. albicans ATCC 32354	3.1	3.1	1.6	0.4	25
C. albicans 83-2-14	12.5	3.1	3.1	0.4	25
C. tropicalis 85-8	12.5	6.3	3.1	0.4	50
Cryptococcus neoformans D49	1.6	0.8	1.6	0.2	0.4
C. neoformans IAM 4514	1.6	1.6	1.6	0.2	0.2
Aspergillus fumigatus IAM 2530	1.6	3.1	1.6	0.4	6.3
A. fumigatus IAM 2034	1.6	3.1	1.6	0.4	6.3
A. flavus FA 21436	6.3	6.3	3.1	6.3	3.1
Fusarium moniliforme A2284	6.3	>100	50	1.6	25
Trichophyton mentagrophytes No. 4329	3.1	3.1	3.1	0.8	0.8
Sporothrix schenckii IFO 8158	0.8	3.1	0.8	0.8	1.6
Petriellidium boydii IFO 8078	6.3	>100	100	6.3	0.8
Mucor spinosus IFO 5317	>100	6.3	100	0.1	100

Table 3. Antifungal spectrum of pradimicins A, B and C by an agar dilution method^a.

^a Yeast morphology agar adjusted to pH 7.0 with 1/15 M phosphate buffer.

Table 4. In vivo antifungal activity of pradimicins A, B and C against systemic infection with Candida albicans A9540 in mice (n = 5).

Table 5. In vivo antifungal activity of pradimicin A against systemic Candida albicans infections with 5-fluorocytosine- and azole-resistant strains in mice (n = 5).

 PD_{50}^{a} (mg/kg/dose, single iv) *C. albicans*

A26090°

5.6

Commound	PD_{50}^{a} (m	ng/kg/dose)
Compound –	Single iv	$Bid \times 2 im^b$
Pradimicin A	7.9	21
Pradimicin B	11	22
Pradimicin C	14	16
Amphotericin B	0.22	0.28
Ketoconazole	44 .	95

^a PD₅₀ value was calculated on day 20 after the fungal challenge.

^b Bid $\times 2$: See a footnote.

Inoculum size: 10LD₅₀.

Amphotericin B0.150.56Ketoconazole> 50> 505-Fluorocytosine> 100> 100

YA22851^b

7.2

^a PD₅₀ value was calculated on day 20 after the fungal challenge.

^b 5-Fluorocytosine-resistant C. albicans.

^e Azole-resistant C. albicans.

Compound

Pradimicin A

Inoculum size: 10LD₅₀.

A9540 infection in mice by iv (single[†]) or im (bid $\times 2^{\dagger}$) administration. The results are summarized in Table 4. Pradimicins A, B and C were active against systemic candidiasis with PD₅₀ values of 7.9 $\sim 14 \text{ mg/kg}$ iv and 16 $\sim 22 \text{ mg/kg/dose}$ im. They were 3 ~ 6 times more potent than ketoconazole but 30 ~ 80 times less potent than amphotericin B. Against 5-fluorocytosine- and azole-resistant *C. albicans* infections, both pradimicin A and amphotericin B were effective at the same dose level by a single iv administration as they were against sensitive *C. albicans* A9540 infection (Table 5). Ketoconazole and 5-fluorocytosine were inactive against both resistant *Candida* strains at 50 and 100 mg/kg iv, respectively.

[†] Treatment schedules of test compounds, single: Once on day 0, bid \times 2: twice a day on days 0 and 1, q2d \times 3: once a day on days 0, 2 and 4, qd \times 5: once a day on days 0 \sim 4 after fungal challenge.

Table 6. In vivo antifungal activity of pradimicin A against systemic Cryptococcus and Aspergillus infections in mice (n = 5).

Compound	PD_{50}^{a} (mg/kg/dose)					
	C. neoform	ans IAM 4514	A. fumigatus IAM 2034			
	Single iv	$Bid \times 2$ im	Single iv	$Bid \times 2$ im		
Pradimicin A	8.9	4.5	21	22		
Amphotericin B	0.30	0.35	0.42	0.20		
Ketoconazole	35	>100	45	84		

 a $\ \mathrm{PD}_{50}$ value was calculated on day 20 after the fungal challenge.

Inoculum size: 10LD₅₀.

Table 7. Antifungal activity of pradimicin A in immunocompromized mic	$e^{a} (n = 5)$).
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			PD ₅₀ ^b (mg/kg/dose	;)	
Compound	C. al	C. albicans		C. neoformans	A. fumigatus
	A9	A9540		IAM 4514	IAM 2034
_	Single iv	Bid $\times 2$ im	(Single iv)	(Single iv) (Single iv)	(Single iv)
Pradimicin A	28	22	7.7	19	45
Amphotericin B	0.56	0.33	0.28	0.56	0.65

^a Mice were pretreated ip with 200 mg/kg of cyclophosphamide 4 days prior to the infection.

 $^{\rm b}~PD_{50}$ value was calculated on day 20 after the fungal challenge.

Inoculum size: 10LD₅₀.

By single iv and bid $\times 2$ im administrations, pradimicin A also showed significant protective effects on systemic infections with *C. neoformans* IAM 4514 and *A. fumigatus* IAM 2034 in mice with PD₅₀ values of 8.9 and 21 mg/kg iv, and 4.5 and 22 mg/kg/dose im, respectively (Table 6). Pradimicin A exhibited good therapeutic efficacy in immunocompromized mice infected systemically with *C. albicans* A9540, *C. tropicalis* CS-07, *C. neoformans* IAM 4514 and *A. fumigatus* IAM 2034 infections

Table 8. Antifungal activity of pradimicin A against Candida and Aspergillus lung infections in mice (n = 5).

	PD_{50}^{a} (mg/kg/dose, q2d × 3 iv)		
Compound	C. albicans A9540	A. fumigatus IAM 2034	
Pradimicin A	18	23	
Amphotericin B	0.71	0.30	

 PD₅₀ value was calculated on day 20 after the fungal challenge.

Inoculum size: 10LD₅₀.

by a single iv dose, the PD₅₀ values being 28, 7.7, 19 and 45 mg/kg, respectively (Table 7). Anti-*C. albicans* activity of pradimicin A was also seen with bid $\times 2$ im treatment.

In Vivo Antifungal Activity against Lung Infections in Mice

Pradimicin A and amphotericin B were tested for therapeutic effects on murine lung infections with C. albicans A9540 and A. fumigatus IAM 2034 by an intravenous $q2d \times 3$ dosing schedule. As summarized in Table 8, both pradimicin A and amphotericin B significantly protected against lethality of mice with lung candidiasis and aspergillosis, the PD₅₀ values of pradimicin A being 18 and 23 mg/kg/dose, respectively.

In Vivo Antifungal Activity against Vaginal and Skin Infections in Mice

The therapeutic efficacy of pradimicin A and amphotericin B was compared against vaginal infec-

Compound	C. albicans A9540 vaginal infection		T. mentagrophytes No. 4329 skin infection	
	ED ₅₀ (mg/kg/dose, iv)	EC ₅₀ (% topical)	EC ₅₀ (% topical)	
Pradimicin A	24	0.33	1.7	
Amphotericin B	0.39	0.10	0.70	

Table 9. Antifungal activity of pradimicin A against Candida vaginal and Trichophyton skin infections in mice (n = 5).

In both iv and topical treatments, test compounds were administered by $qd \times 5$ schedule and their ED₅₀ or EC₅₀ values were calculated on day 7 after the fungal challenge.

tion with *C. albicans* A9540 by qd \times 5 iv and topical treatments in mice. As shown in Table 9, intravenously administered pradimicin A and amphotericin B significantly inhibited vaginal *Candida* infection with PD₅₀ values of 24 and 0.39 mg/kg/ dose, respectively. Both compounds were active but much weaker than ketoconazole by topical treatment. Against skin infection with *T. menta*grophytes No. 4329, topical pradimicin A was also effective with an EC₅₀ value of 1.7%.

Table 10. In vitro cytotoxicity of pradimicin A against various mammalian cells.

Cell	IC ₅₀ (µg/ml)
Vero	> 100
MDCK	>100
HCT-116	> 500
B16-F10	> 500

Medium: Eagle MEM supplemented with FCS (10% for MDCK and B16-F10 cells, and 5% for Vero cells) and antibiotic ($50 \mu g/ml$ of amikacin for MDCK and Vero cells, and $60 \mu g/ml$ of kanamycin for B16-F10 cells), and McCov's 5A medium supplemented with 10% FCS, benzylpenicillin (100 u/ml) and streptomycin (100 $\mu g/ml$) for HCT-116 cells.

Toxicity

In vitro cytotoxicity of pradimicin A was de-

termined against various cultured mammalian cells. As shown in Table 10, pradimicin A was not cytotoxic against all cells tested at 100 or $500 \,\mu$ g/ml. The LD₅₀ values in mice were 120 and more than $400 \,\text{mg/kg}$ by iv and im administrations, respectively. Transient body weight loss was observed in the mice injected at higher doses than 20 and 50 mg/kg by iv and im administrations, respectively.

Antiviral Activity

In vitro antiviral activity of pradimicin A was determined against influenza and herpes simplex viruses by the plaque reduction assay. As shown in Table 11, pradimicin A potently inhibited replication of influenza virus with an IC₅₀ value of $6.8 \,\mu$ g/ml without cytotoxic activity against host MDCK cells at 200 μ g/ml.

Discussion

Pradimicins A, B and C are novel antifungal antibiotics produced by A. hibisca No. P157-2 (ATCC 53557)^{1~7)}. They showed potent *in vitro* antifungal activity and broad antifungal spectrum against a wide variety of fungi and yeasts. Antibacterial activity of pradimicin A was seen against only *Micrococcus luteus* with a MIC value of $3.1 \,\mu\text{g/ml}^{1~3}$. The *in vitro* results suggested their broad therapeutic efficacy in systemic fungal infections, though their *in vitro* potencies were significantly less than that of amphotericin B. In fact, pradimicins A, B and C all exhibited significant protection in the systemic infection with *C. albicans* A9540, which is sensitive to various types of antifungal agents, at similar dose levels by either single iv (PD₅₀= $7.9 \sim 14 \,\text{mg/kg}$) or bid $\times 2 \,\text{im}$ (PD₅₀= $16 \sim 21 \,\text{mg/kg}/\text{dose}$) administration. Pradimicin A was also significantly effective on the systemic infections with azole- or 5-fluorocytosine-resistant *C. albicans*, *A. fumigatus* IAM 2034 and *C. neoformans* IAM 4514 by the same treatment conditions as above. In immunocompromized mice, pradimicin A was active against systemic infections with *C. tropicalis* CS-07

Germand	Influenz	a A virus	Herpes sir	nplex virus
Compound	IC ₅₀ (µg/ml)	TC ₅₀ (µg/ml)	$IC_{50} (\mu g/ml)$	TC_{50} (µg/ml)
Pradimicin A	6.8	> 200	> 200	>200
Ribavirin	1.6	>200	ND	ND
Acyclovir	ND	ND	0.02	>200

Table 11. In vitro antiviral activity of pradimicin A by the plaque reduction assay.

IC₅₀: 50% inhibitory concentration to viral replication.

 TC_{50} : 50% cytotoxic concentration to host cells.

ND: Not done.

as well as the above three pathogens. Pradimicin A was $2 \sim 4$ times less active against *C. albicans, C. neoformans* or *A. fumigatus* in immunocompromized mice than in normal mice by a single iv administration in terms of PD₅₀ values, but its anti-*Candida* activity was equivalent in immunocompromized and normal mice with bid $\times 2$ im treatment. As described above, pradimicin A showed excellent *in vivo* antifungal activity against various systemic fungal infections in both normal and immunocompromized animals. Since, most deep fungal infections are known to be caused by species of *Candida, Aspergillus* and *Cryptococcus neoformans* particularly frequently in immunocompromized patients having cancer, AIDS *etc.*, the above results strongly suggest the clinical usefulness of pradimicin A against a variety of systemic fungal infections.

In the experimental lung infection with *C. albicans* or *A. fumigatus*, and the vaginal infection with *C. albicans*, pradimicin A showed significant therapeutic efficacy by iv administration. Furthermore, the antibiotic inhibited both vaginal and skin infections with *C. albicans* and *T. mentagrophytes*, respectively, with topical application. In vaginal candidiasis, the EC_{50} value of pradimicin A was 0.33%, which is a clinically applicable concentration.

Pradimicin A was not cytotoxic to various cultured mammalian cells at 10 to 100 times higher concentrations $(100 \sim 500 \,\mu\text{g/ml})$ than the antifungal MIC values $(0.8 \sim 12.5 \,\mu\text{g/ml})$. In the acute toxicity experiments in mice, the LD₅₀ values were 120 and more than 400 mg/kg by iv and im administrations, respectively. Pradimicin A was found to possess a Ca⁺⁺-dependent binding capability to yeast cell surfaces but not to mammalian cell membranes⁹). These results suggest that the specific binding to fungal cell surfaces plays, in part, a role in the selective toxicity of pradimicin A against fungal cells. The antibiotic had no mutagenic potential in the Ames test and the *E. coli* reverse mutation assay (unpublished data).

In addition to the antifungal activity, pradimicin A inhibited influenza virus replication in MDCK cells with an IC₅₀ value of $6.8 \,\mu g/\text{ml}$ without cytotoxicity at 200 $\mu g/\text{ml}$. Recently YAMAMOTO *et al.*¹⁰ reported that pradimicin A suppressed the cell damage induced by HIV at concentrations more than $3.5 \,\mu g/\text{ml}$ and suggested its anti-HIV effects on the stage of viral adsorption and cell to cell infection. Taking into consideration the serious systemic fungal infections in AIDS patients, it would be worthwhile to study pradimicin A in the treatment of AIDS patients.

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